

FINAL DRAFT REPORT

Intermedia Transfer Factors for Contaminants Found at Hazardous Waste Sites

1,2 DICHLOROETHANE (DCA)

Risk Science Program (RSP)
Department of Environmental Toxicology
University of California
Davis, California 95616

Prepared for:

The Office of Scientific Affairs
The Department of Toxic Substances Control (DTSC)
and the California Environmental Protection Agency
in Support of the CalTOX Model

December 1994

CONTRIBUTORS

Principal Investigator: Dennis P.H. Hsieh, Sc. D.
Lead Scientist: Thomas E. McKone, Ph. D.
Primary Author: Florence F. Chiao
Authors: Richard C. Currie, B. S.; Florence F. Chiao, Ph. D.; and
Thomas E. McKone, Ph. D.
Information Management: Loreen Kleinschmidt

Contract Manager: Edward Butler, Ph. D.
Department of Toxic Substances Control
301 Capitol Mall, 2nd Floor
Sacramento, CA 95812

TABLE OF CONTENTS

FORWARD.....	i
OVERVIEW.....	ii
CalTOX Chemical-Specific Input Requirements.....	ii
Physicochemical Properties.....	ii
Table 1. Summary of Chemical Properties for 1,2 Dichloroethane.....	iii
The Solid-Water Distribution Coefficients.....	iv
Biotransfer Factors and Bioconcentration Factors.....	iv
Chemical-Specific Transformation Process Half-Lives.....	v
Statistical Methods.....	vi
Mean and Coefficient of Variation.....	vi
Estimation Equations and the Residual Errors of the Estimation Method.....	vii
1,2 Dichloroethane (DCA).....	1
Other Names.....	1
Background.....	1
Formula.....	1
MW: Molecular Weight.....	1
Experimental Values.....	1
K _{ow} : Octanol-Water Partition Coefficient.....	2
Experimental Values.....	2
T _m : Melting Point.....	2
Experimental Values.....	2
VP: Vapor Pressure at Standard Temperatures.....	3
Experimental Values.....	3
Estimation Methods.....	3
Antoine Equation 1.....	3
Antoine Equation 2.....	4
S: Solubility in Water.....	4
Experimental Values.....	4
Unit Conversion.....	4
H: Henry's Law Constant.....	5
Experimental Values.....	5
Estimation Method.....	5
D _{air} : Diffusion Coefficient in Pure Air.....	5
Estimation Method.....	5

D_{water} : Diffusion Coefficient in Pure Water.....	6
Estimation Method.....	6
K_{oc} : Organic-Carbon Partition Coefficient.....	7
Experimental Values.....	7
Estimation Method.....	7
K_{d_s} : Distribution Coefficient in Ground-Surface and Root-Zone Soil.....	8
Estimation Method.....	8
K_{d_v} : Distribution Coefficient in Vadose-Zone Soil.....	8
Estimation Method.....	8
K_{d_q} : Distribution Coefficient in the Ground-Water Zone.....	9
Estimation Method.....	9
K_{d_d} : Distribution Coefficient in Sediment Particles.....	10
Estimation Method.....	10
K_{ps} : Partition Coefficient for Plant-Tissue (Above Ground Fresh Mass) Relative to Soil Concentration (Fresh Soil).....	10
Estimation Method.....	10
K_{pa} : Biotransfer Factors For Plant Leaves Relative to Contaminant Air Concentration.....	11
Estimation Method.....	11
Biotransfer Factors for Food Products.....	12
B_k : Steady-State Biotransfer Factors for Whole Milk Relative to Contaminant Intake by Cattle.....	12
Estimation Method 1.....	12
Estimation Method 2.....	13
B_t : Steady-State Biotransfer Factors for Meat Relative to Contaminant Intake by Cattle.....	14
Estimation Method 1.....	14
Estimation Method 2.....	14
B_e : Steady-State Biotransfer Factors for Egg Relative to Dietary Contaminant Intake by Chickens.....	15
Estimation Method.....	15
B_{bmk} : Biotransfer Factor for Human Breast Milk Relative to Dietary Contaminant Intake by the Mother.....	16
Estimation Method.....	16
BCF : Bioconcentration Factors for Fish Relative to Water Concentration.....	17
Experimental Values:.....	17
Estimation Method.....	17
K_{p_w} : Human Skin Permeability Coefficient Relative to Contaminant Concentration in Water.....	17

Estimation Method.....	18
K_m: Partition Coefficient for Human Skin Relative to Contaminant	
Concentration in Water or Soil.....	18
Estimation Method.....	18
T_{half_a}: Reaction Half-Life in Air.....	19
Experimental Values.....	19
T_{half_g}: Reaction Half-Life in Ground-Surface Soil.....	20
Estimated Values.....	20
T_{half_s}: Reaction Half-Life in Root-Zone Soil.....	20
Estimated Values.....	20
T_{half_v}: Reaction Half-Life in Vadose-Zone Soil.....	21
Experimental Values.....	21
T_{half_q}: Reaction Half-Life in Groundwater.....	22
Experimental Values.....	22
T_{half_w}: Reaction Half-Life in Surface Water.....	22
Experimental Values.....	22
T_{half_d}: Reaction Half-Life in Surface Water Sediment.....	23
Experimental Values.....	23
References.....	23

FORWARD

The Department of Toxic Substances Control (DTSC), within the California Environmental Protection Agency, has the responsibility for managing the State's hazardous-waste program to protect public health and the environment. The Office of Scientific Affairs (OSA) within the DTSC provides scientific assistance in the areas of toxicology, risk, environmental assessment, training, and guidance to the regional offices within DTSC. Part of this assistance and guidance is the preparation of regulations, scientific standards, guidance documents, and recommended procedures for use by regional staff, local governmental agencies, or responsible parties and their contractors in the characterization and mitigation of hazardous-waste-substances-release sites. The CalTOX model has been developed as a spreadsheet model to assist in exposure and health-risk assessments that address contaminated soils and the contamination of adjacent air, surface water, sediments, and ground water.

The modeling effort includes multimedia transport and transformation models, exposure scenario models, and efforts to quantify and reduce uncertainty in multimedia, multiple-pathway exposure models. Use of the CalTOX model requires that we determine the intermedia transfer factors (ITFs) that define concentration relationships between an exposure medium and the environmental medium that is the source of the contaminant. ITFs are chemical and physical parameters which serve as inputs in the CalTOX model analysis.

This report provides a set of ITFs needed to run the CalTOX model for DCA. For this chemical, we have conducted a critical review of existing literature for measured values and estimation methods in order to compute an arithmetic mean (\bar{x}), a coefficient of variation (CV), and plausible range for each ITF.

OVERVIEW

The purpose of this report is to provide a set of chemical-specific intermedia-transfer factors (ITFs) for DCA. We have carried out a critical review of the existing literature in order to identify a mean value, coefficient of variation (CV) and value range for the ITFs listed in Table 1. For values used to define a given parameter, our highest priority was given to experimental values reported in the primary scientific literature, that is, peer-reviewed journals. For parameters that are not readily available from the primary literature, widely cited secondary references such as Lyman et al. (1982, 1990), Verschueren (1984), Howard et al. (1990, 1991), Mackay et al. (1992), the CRC Handbook (1989-90) and the Merck Index (1983, 1989) are used to establish parameter values. When measured values are not available from either the primary literature or secondary references, estimates of ITF parameter values are based on estimation equations that are available in the primary literature. Typically, these estimation methods relate ITFs to other measured contaminant parameters using quantitative-structure-activity-relationship (QSAR) methods. In these cases, parameter values estimated from a QSAR method are treated as the arithmetic mean and the estimation error of the method is used to determine the CV. Table 1 summarizes the units required by the CalTOX model, the values of chemical specific physico-chemical properties, distribution coefficients, biotransfer and bioconcentration factors, and transformation half-lives obtained in this study.

CalTOX Chemical-Specific Input Requirements

The CalTOX model uses three sets of input data—one describing the chemical-specific properties of the contaminants, a second providing properties of the environment or landscape receiving the contaminants, and a third that defines for exposure assessment the characteristics of individuals in various age/sex categories and the characteristics of the micro-environments in which they live or from which they obtain water and food. Each of the inputs in these sets must be described in terms of a mean value with an estimated coefficient of variation, which describes the uncertainty or variability associated with that parameter. This report addresses mean value, CV, and range of values needed to characterize chemical-specific inputs.

Physicochemical Properties

Physicochemical properties include molecular weight, octanol-water partition coefficient, melting point, vapor pressure, Henry's law constant, diffusion coefficients in air and water, and the organic-carbon partition coefficient. The octanol-water partition coefficient provides a measure of the extent of chemical partitioning between water and octanol at equilibrium and is used as a basis for estimating other ITF parameters. The melting point is the temperature at which a compound makes the transition from a solid to a liquid phase. Vapor pressure is the pressure exerted by a chemical vapor in equilibrium with its solid or liquid phase. Water solubility is the upper limit on a chemical's dissolved concentration in pure water, at a specified temperature.

Table 1. Summary of Chemical Properties for 1,2 Dichloroethane

Description	Symbol ^a	Mean Value	Coefficient of Variation	Number of Values
Molecular Weight (g/mol)	MW	98.97	1.9×10^{-4}	4
Octanol-Water Partition Coefficient	K _{ow}	28	0.077	4
Melting Point (K)	T _m	237.7	6.5×10^{-4}	4
Vapor Pressure (Pa)	VP	10,746	0.038	3
Solubility (mol/m ³)	S	86	0.050	4
Henry's Law Constant (Pa·m ³ /mol)	H -	119	0.18	3
Diffusion Coefficient in Pure Air (m ² /d)	D _{air}	0.74	0.05	1
Diffusion Coefficient in Pure Water (m ² /d)	D _{water}	9.7×10^{-5}	0.25	1
Organic Carbon Partition Coefficient	K _{oc} -	18	1.10	2
Distribution Coefficient in Ground-Surface and Root-Zone Soil	K _{d_s} -	b	b	e
Distribution Coefficient in Vadose-Zone Soil	K _{d_v} -	b	b	e
Distribution Coefficient in the Ground-Water Zone	K _{d_q} -	b	b	e
Distribution Coefficient in Ground Water Sediment	K _{d_d} -	b	b	e
Partition Coefficient in Plants Relative to Soil Concentration [ppm (pFM) / ppm (sFM)]	K _{ps} -	1.0	4.0	e
Biotransfer Factor in Plants Relative to Contaminant Air Concentration [m ³ (a)/kg (pFM)]	K _{pa} -	0.015	14	e
Biotransfer Factor in Milk Relative to Cattle-Diet Contaminant Intake (d/kg)	B _k -	5.4×10^{-7}	11	e
Biotransfer Factor in Meat Relative to Cattle-Diet Contaminant Intake (d/kg)	B _t -	6.5×10^{-6}	13	e
Biotransfer Factor in Eggs Relative to Hen-Diet Contaminant Intake (d/kg)	B _e -	2.2×10^{-4}	14	e
Biotransfer in Breast Milk Relative to Contaminant Intake by the Mother (d/kg)	B _{bmk} -	5.6×10^{-6}	10	e
Bioconcentration Factor in Fish Relative to Contaminant Water Concentration	BCF -	2.0	1.0	1
Skin Permeability Coefficient (cm/h)	K _{p_w} -	0.010	2.4	e
Skin-Water/Soil Partition Coefficient [ppm (skin)/ppm (water)]	K _m -	4.2	0.27	e
Reaction Half-Life in Air (d)	T _{half_a}	45	1.2	4
Reaction Half-Life in Ground-Surface Soil (d)	T _{half_g}	110	0.41	4
Reaction Half-Life in Root-Zone Soil (d)	T _{half_s}	110	0.41	4
Reaction Half-Life in the Vadose-Zone Soil (d)	T _{half_v}	260	0.92	6
Reaction Half-Life in Ground-Water Zone Soil (d)	T _{half_q}	335	0.68	5
Reaction Half-Life in Surface Water (d)	T _{half_w}	820	1.6	4
Reaction Half-Life in the Sediment (d)	T _{half_d}	430	0.63	4

^aValues followed by a "-" include default equations that can be used for estimations

^bK_d = [(K_{oc}) × (fraction organic matter)], a site and soil zone specific parameter

^eestimated parameter value

Henry's law constant is a measure at equilibrium of the ratio of chemical activity in the gas above a liquid to chemical activity in the liquid. Diffusion coefficients describe the movement of a molecule in a liquid or gas medium as a result of differences in concentration within the medium. They are used to calculate the dispersive component of chemical transport. The higher the diffusion coefficient, the more likely a chemical is to move in response to concentration gradients. The organic-carbon partition coefficient provides a measure of chemical partitioning between organic carbon (in soils, rocks, and sediments) and water. The higher the K_{OC} the more likely a chemical is to bind to the solid phase of soil or sediment than to the liquid phase.

The Solid-Water Distribution Coefficients

The distribution or sorption coefficient, K_d , is the concentration ratio, at equilibrium, of chemical attached to solids and/or particles (mol/kg) to chemical concentration in the solution, mol/L. When K_{OC} is multiplied by the fraction organic carbon in a soil or sediment, we obtain an estimate of the soil/water or sediment/water partition coefficient. CalTOX requires, as input, distribution coefficients for ground-surface, root-zone, and vadose-zone soil; ground-water-zone rock or soil, and surface-water sediments.

Biotransfer Factors and Bioconcentration Factors

The CalTOX model requires, as input, general relationships that can be used to estimate partition coefficients between air and plants; between soil and plants; between animal feed intake and animal-based food products; between surface water and fish; between the human mother's uptake and breast milk; between skin and water; and between skin uptake and concentration in skin water.

The chemical-specific plant-air partition coefficient, K_{pa} , represents the ratio of contaminant concentration in above-ground plant parts, in mg/kg (fresh mass), to contaminant concentration in the gas-phase of the atmosphere mg/m³ (air). The plant-soil partition coefficient, K_{ps} , expresses the ratio of contaminant concentration in plant parts, both pasture and food, in mg/kg (plant fresh mass) to concentration in wet root-zone soil, in mg/kg.

The biotransfer factors B_t , B_k and B_e are the steady-state contaminant concentrations in, respectively, fresh meat, milk, and eggs; divided by the animals' daily contaminant intake. These factors are expressed in units of (mg/kg)/(mg/d), or kg/d. Unlike bioconcentration factors, which express steady-state concentration ratios between animal tissue and a specific environmental medium, biotransfer factors express the steady-state relationship between intake and tissue or food-product concentrations.

Lactating women can transfer to breast milk their intake of contaminants from all intake routes—ingestion, inhalation, and dermal contact. B_{bmk} is the biotransfer factor for milk-concentration versus the mother's intake. This relationship may also be

described as the ratio of contaminant concentration in mother's milk divided by the mother's daily intake of that contaminant, in units of d/kg (milk).

The bioconcentration factor BCF provides a measure of chemical partitioning between fish tissue based on chemical concentration in water.

Chemical specific exposure factors used in CalTOX include the skin-water and skin-soil partition coefficients. K_m is the skin-water partition coefficient in cm^3 (water)/ cm^3 (skin) . In order to estimate the skin-soil partition factor, K_m^{soil} , with units $\text{cm}^3(\text{soil})/\text{cm}^3(\text{skin})$, we divide equation K_m by the sorption coefficient K_d for soil, or

$$K_m^{\text{soil}} = \frac{K_m}{K_d}$$

K_{p_w} is the steady-state permeability coefficient in cm/hour for a contaminant from water on skin through stratum corneum and can either be based on a measured value or estimated values.

Chemical-Specific Transformation Process Half-Lives

Chemical transformations, which may occur as a result of biotic or abiotic processes, can have a profound effect on the persistence of contaminants in the environment. Experimental methods and estimation methods are available for defining these fate processes in a variety of media. Specific information on the rates and pathways of transformation for individual chemicals of concern should be obtained directly from experimental determinations, if possible, or derived indirectly from information on chemicals that are structurally similar. CalTOX makes use of media- and reaction-specific reaction half-lives to establish rate constants for transformation removal processes that include photolysis, hydrolysis, oxidation/reduction, and microbial degradation.

Transformation-rate half-lives are among the more uncertain parameters in the CalTOX model. There are typically few available measurements or ranges of estimated values in the primary and secondary literature. Most of the available half-life values are obtained from limited measurements for environmental media that are not necessarily representative of those in California. These values often involve scientific judgment as much as measurement. In making use of these data, we expanded the range of the reported values by a factor of 5 when only 2 or 3 representative values are presented and by a factor of 10 when only one value is provided. If 4 or more measured values are available, these uncertainty factors are not applied. In order to express the lack of reliability associated with a limited number of measured values for a parameter, these uncertainty factors are used to express both large uncertainty and significant variability.

Statistical Methods

Each of the inputs to CalTOX must be described by a mean value and an estimated coefficient of variation which describes the uncertainty or variability associated with that parameter. For input values that are derived from a number of measured values, the mean and coefficient of variation are obtained from the arithmetic mean and the arithmetic standard deviation of the inputs. For estimated input values, the mean and coefficient of variation are obtained from an estimation equation and the residual error of the estimation equation. The methods we used to obtain these values are described here.

Mean and Coefficient of Variation

The arithmetic mean (\bar{x}) is used to represent all inputs that are derived from a number of measured values—even those that might have geometric distributions. The (\bar{x}) is computed by summing the reported values and dividing this sum by the total number of observations:

$$\text{Arithmetic mean } (\bar{x}) = \frac{\sum_{i=1}^n x_i}{n} \quad (\text{Eqn. 1})$$

Where $\sum_{i=1}^n x_i$ is the sum of the observed values and n is the number of observations. In this case, the coefficient of variation (CV) is computed by dividing the arithmetic standard deviation (S_n) by the mean. Standard deviation and CV are computed according to the following equations:

$$\text{standard deviation } (S_n) = \sqrt{\frac{\sum_{i=1}^n (x_i - \bar{x})^2}{n}} \quad (\text{Eqn. 2})$$

$$\text{coefficient of variation } (CV) = \frac{S_n}{\bar{x}} \quad (\text{Eqn. 3})$$

It should be noted that, based on the central limit theorem of statistics, the confidence associated with the estimate of \bar{x} from above becomes large as the number of samples used to estimate \bar{x} also becomes large. Therefore, the reliability of the estimates of *mean* and *CV* of a parameter are low when the sample size is small. It is beyond the scope of this document to explicitly address the reliability of these estimates. Nonetheless, in order to give an indication of potential reliability problems, we list the number of measurements used to estimate the mean and CV of each parameter in the last column of Table 1.

Estimation Equations and the Residual Errors of the Estimation Method

Estimates of some CalTOX inputs are based on regression equations that relate a parameter value to some measure of structure or activity associated with the contaminant. These methods are referred to as quantitative structure-activity relationship (QSAR) methods. The reliability of a parameter-value estimated in this way is defined by the precision of these QSAR methods.

Our estimate of precision in QSAR estimation methods is based on calculating, S_e , the *standard error of the estimate* (or standard deviation of the residuals). This error calculation is based on the regression equations and fragment models used to derive a parameter value. To illustrate, when the value of parameter such as the organic-carbon partition coefficient (K_{oc}) is estimated using a regression or correlation analysis, the S_e is calculated using the following approach (Hamburg, 1970). First, since it is typical that it is the $\log K_{oc}$ (not K_{oc} itself) that is estimated from a regression equation, we calculate the S_e of $\log K_{oc}$ according to

$$S_e \text{ of } \log K_{oc}^{est} = \sqrt{\frac{\sum_{i=1}^n (\log K_{oc}^{msd} - \log K_{oc}^{est})^2}{(n-2)}} \quad (\text{Eqn. 4})$$

where n is the number of chemicals used in the estimation protocol and K_{oc}^{est} refers to the estimated property (K_{oc} in this case) and K_{oc}^{msd} refers to the corresponding measured values used to carry out the regression. In order to calculate the S_e of K_{oc} we make use of the transformation

$$\text{GSD} (K_{oc}^{est}) = 10^{(S_e \text{ of } \log K_{oc}^{est})} \quad (\text{Eqn. 5})$$

to calculate the geometric standard deviation of S_e (GSD) of K_{oc}^{est} , which is simply the GSD of the K_{oc} estimate, that is $\text{GSD} (K_{oc}^{est})$. It has been shown by Atchison and Brown (1957) that the relationships between the GSD and CV for log normal distributions are as follows

$$\text{GSD} = \exp\{\sqrt{\ln(1+CV^2)}\} \quad (\text{Eqn. 6})$$

$$CV = \sqrt{\left(\exp\{[\ln(\text{GSD})]^2\} - 1\right)} \quad (\text{Eqn. 7})$$

Since the implicit assumption of a regression for estimating the log of K_{oc} is that any estimated value, $\log(K_{oc}^{est})$, is centered on normal distribution with standard deviation equal to S_e of $\log K_{oc}$ it follows that the corresponding estimated value of K_{oc} is centered on a log normal distribution with GSD (K_{oc}^{est}) and with

$$CV(K_{oc}^{est}) = \sqrt{\exp\{[\ln(\text{GSD}(K_{oc}^{est}))]^2\} - 1} \quad (\text{Eqn. 8})$$

This approach is used to estimate CVs for the estimation equations presented in this document.

In some cases the error term, CV for example, is calculated by combining through the operations of multiplication and division the CVs of two or more parameters. For example the CV in the ration $H = VP/S$ is combined from the CV (VP) and CV (S). In this case, if the input parameters are independent, the combined CV is calculated using the following equation:

$$CV_{\text{combined}} = \sqrt{\frac{\sum_{i=1}^n CV_i^2}{n}} \quad (\text{Eqn. 9})$$

where n is the number of parameters used in the multiplication/division and CV_i is the coefficient of variation in the i th input parameter.

1,2 Dichloroethane (DCA)

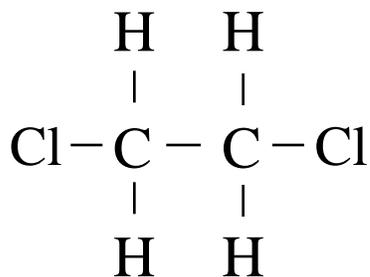
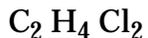
Other Names

1,2 DCA, DCE, DCA, EDC, dichloroethylene, sym-dichloroethane; glycol dichloride; 1,2 bichloroethane; , -dichloroethane; ENT 1,656; ethane dichloride; ethylene chloride; 1,2 ethylene dichloride; Dichloroemulsion; Borer Sol; Dutch Liquid; Dutch Oil; Brocide; RCRA Waste Number U077; CAS 107-06-2 (WHO IARC, 1979)

Background

1,2 Dichloroethane (DCA) belongs to the chemical family of chlorinated alkanes. DCA is manufactured by reacting ethylene with chlorine or hydrogen chloride in the presence of a metal catalyst. Used in the manufacturing of vinyl chloride, tetraethyllead, intermediate insecticidal fumigants and tobacco flavoring; DCA is also found as a constituent in paint, varnish, finish removers, soaps and scouring compounds; as a metal degreaser; in wetting and penetrating agents, chemical synthesis and ore floatation (Verscheuren, 1983). It is also used in the production of ethyleneamines, tetrachloroethylene; trichloroethylene, as a lead scavenger and in the production of vinylidene chloride (WHO IARC, 1979). Disposal of EDC tars is usually by burial in a landfill or incineration. The lack of detection of DCA in soil is due to its rapid removal into water and air (Scherb, 1978). Evaporation appears to be the major pathway by which DCA is lost from the water (Dilling 1975). In the atmosphere, oxidation by hydroxyl radicals is the dominant loss process (Radding, 1977).

Formula



MW: Molecular Weight

The units used for molecular weight are grams/mole (g/mol).

Experimental Values

- | | |
|-------|--|
| 98.96 | reported in Weast et al. [CRC Handbook (1990)] |
| 98.96 | reported in Budavari et al. [Merck Index (1989)] |

- 98.97 reported in Patty's Industrial Hygiene & Toxicology [Clayton and Clayton (1981)]
- 99 reported in Verscheuren (1983)

From the above 4 reported values above, we obtain the following statistics for the molecular weight of 1,2 DCA:

Arithmetic mean (coefficient of variation):
 MW = 98.97 (1.9×10^{-4}) g/mol

Range: 98.96 to 99 g/mol

K_{ow}: Octanol-Water Partition Coefficient

The units used for K_{ow} are $\frac{\text{mg/liter (octanol)}}{\text{mg/liter (water)}}$ and K_{ow} is therefore unitless.

Experimental Values

- 25 reported at 20 °C as a K_{ow} of 25 by Chiou et al. (1977) using a shake flask-GC method
- 28.2 reported as a log K_{ow} of 1.45 by Veith et al. (1980) estimated using reverse-phase liquid chromatography (RP-HPLC)
- 28.2 reported at 23 °C as a K_{ow} of 28.2 by Banerjee et al. (1980) using a shake-flask LSC method [Also cited in Mackay et al.(1993)]
- 30.2 reported as a log K_{ow} of 1.48 by Jow and Hansch (1985) as unpublished results in Hansch and Leo (1979)

From the 4 measured values above we obtain the following statistics for the octanol-water partition coefficient of 1,2 DCA at 25 °C:

Arithmetic mean (coefficient of variation):
 K_{ow} = 28 (0.077)

Range: 25 to 30.2

T_m: Melting Point

The units used for melting point are kelvins (K).

Experimental Values

- 237.49 reported as a MP of -35.66 °C by Thermodynamics Research Center Data Project (1964) [also cited in Riddick et al. (1986)]

- 237.7 reported as a MP of -35.5 °C by McGovern (1943)
 237.8 reported as a MP of -35.4 °C by Verscheuren (1983)
 237.8 reported as a MP of -35.3 °C by Weast et al. [CRC (1988)]

From the 4 measured values above, we obtain the following statistics for the melting point of 1,2 DCA:

Arithmetic mean (coefficient of variation):

$$T_m = 237.7 (6.5 \times 10^{-4}) \text{ K}$$

Range: 237.49 to 237.8 K

VP: Vapor Pressure at Standard Temperatures

The units used for vapor pressure are pascals (Pa).

Experimental Values

- 10,399 reported at 25 °C as 78 mm Hg by McGovern (1943)
 10,639 reported at 25 °C by Pearce and Peters (1929) [also cited in Boublik et al. (1984)]
 11,199 reported at 25 °C as 84 mm Hg extrapolated from Stull (1947)

From the 3 measured values above, we obtain the following statistics for the vapor pressure of 1,2 DCA at 25 °C:

Arithmetic mean (coefficient of variation):

$$\text{VP} = 10,700 (0.038) \text{ Pa}$$

Range: 10,399 to 11,199 Pa

Estimation Methods

Antoine Equation 1

The following Antoine equation was derived from data by Pearce and Peters (1929) [also cited in Boublik et al. (1984)].

$$\log_{10} \text{VP} = 6.16284 - \frac{1278.323}{223.694 + T (\text{°C})}$$

for -30 °C < T < 99 °C; yielding:

$$\text{VP} = 10,500 \text{ Pa at } 298 \text{ K}$$

Antoine Equation 2

The following Antoine equation was derived from data by Stull (1947).

$$\log_{10} VP = 10.1193 - \frac{1813.12}{273 + T (\text{°C})}$$

for $-24 \text{ °C} < T < 64 \text{ °C}$; yielding:

$$VP = 10,800 \text{ Pa at } 298 \text{ K}$$

S: Solubility in Water

The units used in the solubility values below are $\frac{\text{mg}}{\text{liter (water)}} \text{ (mg/L)}$.

Experimental Values

7987	reported at 25 °C as 80700 μM by Banerjee et al. (1980) using a shake-flask GC method [Also cited in Horvath (1982)]
8450	reported at 20 °C as 0.845 g/100 ml solution by Chiou et al. (1977) using a shake flask-GC method
8650	reported at 25 °C as 0.865 g/100 g water by Gross (1929) using a shake-flask interferometer method [also cited in MacKay et al. (1990)]
9000	reported as 9.0 g/1000 g water by Gross and Saylor (1931) using a shake-flask interferometer method

Unit Conversion

$$\begin{aligned} &\text{Arithmetic mean (coefficient of variation) of 1,2 DCA solubility} \\ &= 8522 (0.050) \text{ mg/L} \\ &= 86 (0.050) \text{ mol/m}^3 \end{aligned}$$

From the 4 measured values above, we obtain the following statistics for the water solubility of 1,2 DCA at 25 °C:

$$\begin{aligned} &\text{Arithmetic mean (coefficient of variation):} \\ &S = 86 (0.050) \text{ mol/m}^3 \end{aligned}$$

$$\text{Range: } 80.7 \text{ to } 90.9 \text{ mol/m}^3$$

H: Henry's Law Constant

The units used for Henry's Law constant are $\frac{\text{Pascals}\cdot\text{m}^3}{\text{mole}}$ (Pa·m³/mol).

Experimental Values

- 101 reported at 20 °C as 0.0010 atm m³/mol by Tse (1992)
- 111 reported at 25 °C as 0.0011 atm m³/mol by EPA (1987) from a batch stripping method
- 143 reported as 0.00141 atm m³/mol at 25 °C by Ashworth et al. (1988) measured by EPICS [Also cited in Mackay et al. (1992)]

From the 3 measured values above, we obtain the following statistics for Henry's law constant at 20-25 °C:

Arithmetic mean (coefficient of variation):
H = 120 (0.18) Pa·m³/mol

Range: 101 to 143 Pa·m³/mol

Estimation Method

$$H = \frac{VP \text{ (Pa)}}{S \text{ (mol/m}^3\text{)}} = \frac{10700}{86} = 125 \text{ Pa}\cdot\text{m}^3/\text{mol}$$

D_{air}: Diffusion Coefficient in Pure Air

The units used for the diffusion coefficient in pure air are $\frac{\text{meters}^2}{\text{day}}$ (m²/d).

Estimation Method

Based on the Fuller et al. (1966) method described in Lyman et al. (1982), the estimated diffusion coefficient in air(m²/d) is given by:

$$D_{\text{air}} = 8.6 \times 10^{-3} T^{1.75} \frac{\sqrt{(29 + M_x)/(29 \times M_x)}}{\left[2.7 + V_x^{1/3}\right]^2}$$

Molar volume(V_x) can be estimated by the LeBas incremental method as described in Lyman et al. (1982) With a molar volume, V_x, of 94 cm³/mol, molecular weight (M_x) of 96.97 g/mol, and a temperature equal to 298 K, the above expression gives:

$$D_{\text{air}} = 3.46 \times 10^{-5} T^{1.75} = 0.74 \text{ m}^2/\text{d}$$

The reported average absolute estimation error is 5 to 10% [Fuller et al. (1966)]. This estimation error is reported as <5% for chlorinated aliphatics and equivalent to the CV below.

Based on the estimated value and the estimation error reported above, we obtain the following statistics for the estimated air diffusion coefficient of 1,2 DCA at 25 °C:

Arithmetic mean (coefficient of variation):

$$D_{\text{air}} = 0.74 (0.05) \text{ m}^2/\text{d}$$

D_{water}: Diffusion Coefficient in Pure Water

The units used for the diffusion coefficient in pure water are $\frac{\text{meters}^2}{\text{day}}$ (m²/d).

Estimation Method

Based on the Wilke and Chang (1955) method described in Reid et al. (1987) the diffusion coefficient in water (m²/d) is given by:

$$D_{\text{water}} = \frac{6.5 \times 10^{-7} \sqrt{f \times M_y} T}{h_y V_x^{0.6}}$$

Wilke and Chang (1955) recommend an association factor, *f*, of 2.6 when the solvent is water. The viscosity of water, *h_y*, is 0.89 cP at 25 °C. Molar volume (*V_x*) can be estimated by the LeBas incremental method as described in Lyman et al. (1982). With a *V_x* equal to 94 cm³/mol, a temperature (*T*) of 298 K, and *M_y* (MW of water) equal to 18 g/mol., this expression gives:

$$D_{\text{water}} = 3.27 \times 10^{-7} T = 9.7 \times 10^{-5} \text{ m}^2/\text{d at } 25 \text{ }^\circ\text{C}$$

Original data for this estimation, provided in Reid et al. (1987), can be used to determine the standard error of the estimator for this estimation method. From this data we calculate a CV of 0.25 from a 25% estimation error.

Based on the estimated value and the estimation error reported above, we obtain the following statistics for the estimated water diffusion coefficient of 1,2 DCA at 25 °C:

Arithmetic mean (coefficient of variation):

$$D_{\text{water}} = 9.7 \times 10^{-5} (0.25) \text{ m}^2/\text{d}$$

K_{oc}: Organic-Carbon Partition Coefficient

The units used for K_{oc} are $\frac{\text{mg/kg (organic carbon)}}{\text{mg/kg (water)}}$ and K_{oc} is therefore unitless.

Experimental Values

- 4.1 reported at 25 °C as a K_{oc} of 2.1 to 6.1 by Liljestrand and Charbeneau (1987) using an organic clay soil from an aquifer (19 ft.) with a fraction organic carbon (f_{oc}) of 0.169 from an aquifer
- 33 reported at 25 °C as a K_d of 0.304 by Chiou et al. (1979) using a Willamette silt loam (pH = 6.8; f_{om} = 1.6%) and assuming that organic carbon is 58% of organic matter (Schwartzbach et al. (1993) [Also cited in Mackay et al. (1993)]

From the 2 measured values above, we obtain the following statistics for the organic carbon partition coefficient for 1,2 DCA:

Arithmetic mean (coefficient of variation):

$$K_{\text{oc}} = 18 (1.1)$$

Range: 4.1 to 33

Estimation Method

Karickhoff (1981) has described empirical estimation methods for obtaining K_{oc} from K_{ow}. The most general of these is that K_{oc} is equal to 0.41 times K_{ow}.

$$K_{\text{oc}} = 0.41 \times K_{\text{ow}}$$

$$K_{\text{ow}} = 28$$

$$K_{\text{oc}} (\text{est}) = 11 (1)$$

The reported CV is based on data provided by Karickhoff (1981). This estimation error does not include uncertainty in the value of K_{ow}.

K_{d_s}: Distribution Coefficient in Ground-Surface and Root-Zone Soil

The units used for K_{d_s} are $\frac{\text{mg/kg (dry surface and root-zone soil)}}{\text{mg/kg (water)}}$ and K_{d_s} is therefore unitless.

Estimation Method

This is a site specific parameter and depends on the fraction organic carbon in the surface and root-zone soil and on the value of K_{oc}. K_{d_s} is the product of the soil organic carbon partition coefficient (K_{oc}) and the fraction organic carbon in the surface and root-zone soil (f_{oc_s}) (Karickhoff, 1981).

$$K_{d_s} = K_{oc} \times f_{oc_s}$$

$$f_{oc_s} = \frac{\text{kg organic carbon (dry surface and root-zone soil)}}{\text{kg (soil)}}$$

Based on the estimation reported above, we obtain the following equation for the distribution coefficient in surface and root-zone soil. K_{d_s} is a site and soil-zone specific parameter depending on the fraction organic carbon in the surface and root-zone soil or:

$$K_{d_s} = K_{oc} \times f_{oc_s}$$

K_{d_v}: Distribution Coefficient in Vadose-Zone Soil

The units used for K_{d_v} are $\frac{\text{mg/kg (dry vadose-zone soil)}}{\text{mg/kg (water)}}$ and K_{d_v} is therefore unitless.

Estimation Method

This is a site specific parameter and depends on the fraction organic carbon in the vadose-zone soil and on the value of K_{oc}. K_{d_v} is the product of the soil organic carbon partition coefficient (K_{oc}) and the fraction organic carbon in the vadose-zone soil (f_{oc_v}) (Karickhoff, 1981).

$$K_{d_v} = K_{oc} \times f_{oc_v}$$

$$f_{oc_v} = \frac{\text{kg organic carbon (dry vadose-zone soil)}}{\text{kg (soil)}}$$

Based on the estimation reported above, we obtain the following equation for the distribution coefficient in vadose-zone soil. K_{d_v} is a site and soil-zone specific parameter depending on the fraction organic carbon in the vadose-zone or:

$$K_{d_v} = K_{oc} \times f_{oc_v}$$

K_{d_q} : Distribution Coefficient in the Ground-Water Zone

The units used for K_{d_q} are $\frac{\text{mg/kg (dry aquifer material)}}{\text{mg/kg (water)}}$ and K_{d_q} is therefore unitless.

Estimation Method

This is a site-specific parameter and depends on the fraction organic carbon in the ground-water zone and on the value of K_{oc} . K_{d_q} is the product of the soil organic carbon partition coefficient (K_{oc}) and the fraction organic carbon in the ground-water zone (f_{oc_q}) (Karickhoff, 1981).

$$K_{d_q} = K_{oc} \times f_{oc_q}$$

$$f_{oc_q} = \frac{\text{kg organic carbon (dry aquifer material)}}{\text{kg (solid)}}$$

Based on the estimation reported above, we obtain the following equation for the distribution coefficient in the ground-water zone. K_{d_q} is a site and soil-zone specific parameter depending on the fraction organic carbon in the ground-water zone or:

$$K_{d_q} = K_{oc} \times f_{oc_q}$$

K_{d_d} : Distribution Coefficient in Sediment Particles

The units used for K_{d_d} are $\frac{\text{mg/kg (dry surface-water sediment)}}{\text{mg/kg (water)}}$ and K_{d_d} is therefore unitless.

Estimation Method

This is a site specific parameter and depends on the fraction organic carbon in the surface-water sediment and the value of K_{oc} . K_{d_d} is the product of the soil organic carbon partition coefficient (K_{oc}) and the fraction of organic carbon in surface-water sediment (f_{oc_d}) [Karickhoff, 1981].

$$K_{d_d} = K_{oc} \times f_{oc_d}$$

$$f_{oc_d} = \frac{\text{kg organic carbon (dry surface-water sediment)}}{\text{kg (soil)}}$$

Based on the estimation reported above, we obtain the following equation for the distribution coefficient in surface-water sediment particles. K_{d_d} is a site and soil-zone specific parameter depending on the fraction organic carbon in surface-water sediment or:

$$K_{d_d} = K_{oc} \times f_{oc_d}$$

 K_{ps} : Partition Coefficient for Plant-Tissue (Above Ground Fresh Mass) Relative to Soil Concentration (Fresh Soil)

The units used for K_{ps} are $\frac{\text{mg/kg (plant fresh mass [pFM])}}{\text{mg/kg (soil fresh mass [sFM])}}$ (ppm [pFM]/ppm [sFM]).

No reported measurements of K_{ps} for 1,2 DCA are available in the current literature. An estimation method for this parameter is thus applied.

Estimation Method

Based on a review of reported measurements of bioconcentration for 29 persistent organochlorines in plants, Travis and Arms (1988) have correlated plant-soil bioconcentration (on a dry-mass basis) in above-ground plant parts with octanol-water partition coefficients. This bioconcentration factor, B_v , on a dry-weight basis is expressed as:

$$\log B_v = 1.58 - 0.58 \log K_{ow} \pm 0.73 \quad (n=29, r^2=0.525)$$

We calculated the error term, ± 0.73 , from the mean square error of the estimator for this regression from the data provided by Travis and Arms (1988). When adjusted to a fresh-mass basis (assuming that the plant dry-mass fraction equals 0.2), this estimation equation gives the plant-soil partition coefficient, K_{ps} , expressing the ratio of contaminant concentration in mg/kg in above-ground plant fresh mass relative to contaminant concentration in mg/kg (dry soil) in the root-zone as:

$$K_{ps} = 7.7 K_{ow}^{-0.58} (CV = 4.0) \text{ ppm (pFM)/ppm (sDM)}$$

Expressing the ratio of contaminant concentration in mg/kg in above-ground plant fresh mass relative to contaminant concentration in mg/kg (fresh soil), and assuming fresh soil 10% by mass water in the root-zone soil, the K_{ps} estimation is:

$$K_{ps} (\text{est}) = 7.0 K_{ow}^{-0.58}$$

$$K_{ow} = 28$$

$$K_{ps} (\text{est}) = 1.0 \text{ ppm (pFM)/ppm (sFM)}$$

The estimation error reported above corresponds to a CV of 4.

From the estimation method identified above, we obtain the following statistics for the partition coefficient in plant leaves relative to contaminant concentration in soil of 1,2 DCA:

Arithmetic mean (coefficient of variation):

$$K_{ps} = 1.0 (4) \text{ ppm (pFM)/ppm (sFM)}$$

K_{pa} : Biotransfer Factors For Plant Leaves Relative to Contaminant Air Concentration

The units used for K_{pa} are $\frac{\text{mg/kg (plant fresh mass [pFM])}}{\text{mg/cubic meter of air (m}^3 \text{ [air])}} (\text{m}^3 \text{ [a]}/\text{kg [pFM]})$

No reported measurements of K_{pa} for 1,2 DCA are available in the current literature. An estimation method for this parameter is thus applied.

Estimation Method

Based on the model of Riederer (1990) for foliar uptake of gas-phase contaminants (mg/m^3) relative to contaminant concentration in plant leaves (mg/kg fresh mass), we estimate a steady-state plant-air coefficient as:

$$K_{pa} (\text{m}^3 [\text{a}]/\text{kg} [\text{pFM}]) = [0.5 + ((0.4 + 0.01 \times K_{ow})(RT/H))] \times 10^{-3} \text{ kg}/\text{m}^3$$

$$R = 8.313 \text{ Pa}\cdot\text{m}^3/\text{mol}\cdot\text{K}$$

$$T = 298 \text{ K}$$

$$H = 120 \text{ Pa}\cdot\text{m}^3/\text{mol}$$

$$K_{ow} = 28$$

$$K_{pa} (\text{est}) = 0.015 \text{ m}^3 [\text{a}]/\text{kg} [\text{pFM}]$$

McKone (1993) has estimated that the CV associated with this partition estimation model is on the order of 14.

From the estimation method identified above, we obtain the following statistics for the partition coefficient in plant leaves relative to contaminant concentration in air for 1,2 DCA:

Arithmetic mean (coefficient of variation):

$$K_{pa} = 0.015 (14) \text{ m}^3 [\text{a}]/\text{kg} [\text{pFM}]$$

BIOTRANSFER FACTORS FOR FOOD PRODUCTS

The biotransfer factors B_t , B_k and B_e are the steady-state contaminant concentrations in, respectively; fresh meat, milk, and eggs; divided by the animals daily contaminant intake, and are expressed in units of (mg/kg)/(mg/d) or kg/d.

B_k : Steady-State Biotransfer Factors for Whole Milk Relative to Contaminant Intake by Cattle

The units used for B_k are days/kg (milk) (d/kg [milk]).

No reported measurements of B_k are available in the current literature.

Estimation methods are therefore considered.

Estimation Method 1

Based on a review of biotransfer factors for 28 organic chemicals in milk Travis and Arms (1988) developed the following geometric-mean regressions for B_{k1} based on the octanol-water partition coefficient, K_{ow} ,

$$\log B_{k1} = \log K_{ow} - 8.1 \pm 0.84 \quad (n = 28, r^2 = 0.55)$$

Using the data provided by Travis and Arms (1988), we calculated the error term, ± 0.84 , from the mean square error of the estimator for this regression. This estimation error corresponds to a CV of 6. From the above expression and $\log K_{ow}$ of 1.45, we obtain the following statistics for the B_{k1} of 1,2 DCA:

$$B_{k1} \text{ (est)} = 2.2 \times 10^{-7} \text{ days/kg (milk)}$$

$$CV = 6$$

Estimation Method 2

The transfer of organic chemicals from feed to milk has also been expressed in terms of the fat-diet partition coefficient, K_{fd} , which is the steady-state ratio of contaminant concentration in animal fat (or lipid) to contaminant concentration in animal feed with units kg (feed)/kg (fat). Kenaga (1980) reviewed cattle-dietary feeding studies for 23 chemicals, and from these studies derived the following fat-feed equation relating K_{fd} to K_{ow} ,

$$\log K_{fd} = 0.5 \log K_{ow} - 3.457 \pm 1 \text{ (n = 23, } r^2 = 0.62)$$

The estimation error in this expression, ± 1 , was calculated by Kenaga (1980). From the above expression with $\log K_{ow}$ of 1.45, an assumed pasture intake by dairy cattle of 85 kg/d (McKone and Ryan, 1989), and an assumed fat content of 0.04 in milk; we obtain the following statistics for the B_{k2} of 1,2 DCA:

$$B_{k2} \text{ (est)} = 8.7 \times 10^{-7} \text{ days/kg (milk)}$$

$$CV = 14$$

The above estimation error corresponds to assumed CV of 14.

The 2 estimation values reported above yield the arithmetic mean and CV reported below:

$$B_k \text{ (avg)} = 5.5 \times 10^{-7} \text{ days/kg (milk)}$$

$$CV_{\text{pooled}} = 11$$

Based on the estimation equation and the estimation error reported above, we obtain the following value for the estimated steady-state biotransfer factor for milk relative to dietary contaminant intake by dairy cattle for 1,2 DCA:

Arithmetic mean (coefficient of variation):
 $B_k = 5.5 \times 10^{-7} \text{ (11) days/kg (milk)}$

Range: 2.2×10^{-7} to 5.5×10^{-7} days/kg (milk)

B_t: Steady-State Biotransfer Factors for Meat Relative to Contaminant Intake by Cattle

The units used for B_t are days/kg (meat) (d/kg [meat]).

No reported measurements of cattle-meat biotransfer for 1,2 DCA are available in the current literature. Estimation methods are therefore considered.

Estimation Method 1

Based on a review of biotransfer factors for 36 chemicals in meat, Travis and Arms (1988) developed the following geometric-mean regression for B_{t1} based on the octanol-water partition coefficient, K_{ow},

$$\log B_{t1} = \log K_{ow} - 7.6 \pm 0.95 \quad (n = 36, r^2 = 0.67)$$

Using the data provided by Travis and Arms (1988), we calculated the error term, ± 0.95 from the mean square error of the estimator for this regression. This estimation errors corresponds to a CV of 11. From the above expression and a log K_{ow} equal to 1.45, we obtain the following estimation:

$$B_{t1} \text{ (est)} = 7.0 \times 10^{-7} \text{ days/kg (meat)}$$

$$CV = 11$$

Estimation Method 2

The transfer of organic chemicals from feed to meat has also been expressed in terms of the fat-feed partition coefficient, K_{fd}, which is the steady-state ratio of contaminant concentration in animal fat (or lipid) to contaminant concentration in animal feed with units kg(feed)/kg(fat). Kenaga (1980) reviewed cattle-dietary feeding studies for 23 chemicals, and from these studies derived the following fat-feed equation relating K_{fd} to K_{ow}:

$$\log K_{fd} = 0.5 \log K_{ow} - 3.457 \pm 1 \quad (n = 23, r^2 = 0.62)$$

The estimation error in this expression, ± 1 , was calculated by Kenaga (1980). From the above expression with log K_{ow} equal to 1.45, an assumed pasture intake by beef cattle of 60 kg/d (McKone and Ryan, 1989), and an assumed fat content of 0.4 in meat; we obtain the following estimation:

$$B_{t2} \text{ (est)} = 1.2 \times 10^{-5} \text{ days/kg (meat)}$$

$$CV = 14$$

The above estimation error corresponds to a CV of 14.

The 2 estimation values reported above yield the arithmetic mean and CV reported below:

$$B_t \text{ (avg)} = 6.5 \times 10^{-6} \text{ days/kg (meat)}$$

$$CV_{\text{pooled}} = 13$$

Based on the estimation equation and the estimation error reported above, we obtain the following value for the estimated steady-state biotransfer factor for meat relative to dietary contaminant intake by cattle for 1,2 DCA:

Arithmetic mean (coefficient of variation):

$$B_t = 6.5 \times 10^{-6} \text{ (13) days/kg (meat)}$$

Range: 7.0×10^{-7} to 1.2×10^{-5} days/kg (meat)

B_e : Steady-State Biotransfer Factors for Egg Relative to Dietary Contaminant Intake by Chickens

The units used for B_e are days/kg (eggs) (d/kg [eggs]).

No reported measurements of egg-feed biotransfer for 1,2 DCA are available in the current literature. An estimation method is therefore considered.

Estimation Method

Based on measurements of polychlorodibenzodioxins (PCDDs) and polychlorodibenzo-furans (PCDFs) concentrations in soil versus concentrations in egg-fat and adipose tissue of foraging chickens, Stephens et al. (1990) have shown that contaminant concentrations in animal fat correlate with soil concentrations. In addition, they found the fat-soil partition factor in chicken fat is roughly six times higher than the fat-soil partition factor in cattle. However, the fraction of total intake represented by soil in the chicken feed is higher than in the cattle feed. Based on these observation and what is discussed in the above B_k and B_t sections, we (a) assume that the fat-feed partition factor in chickens is similar to that in cattle, (b) use $\log K_{fd} = \log K_{ow} - 4.9$ to estimate the K_{fd} for chickens, and (c) use the fat content of eggs (0.08) and feed intake of chickens (0.12 kg/d [fresh mass]) to obtain the following estimate of a biotransfer factor, B_e , from chicken feed to eggs with units d/kg(eggs):

$$\log B_e = \log K_{ow} - 5.1$$

$$\log K_{ow} = 1.45$$

$$B_e = 2.2 \times 10^{-4} \text{ d/kg (eggs)}$$

We estimate the CV in this expression is 14.

Based on the estimation equation and the estimation error reported above, we obtain the following value for the estimated steady-state biotransfer factors for egg concentration relative to dietary contaminant intake by chickens for 1,2 DCA:

Arithmetic mean (coefficient of variation):
 $B_e = 2.2 \times 10^{-4}$ (14) days/kg (eggs)

B_{bmk} : Biotransfer Factor for Human Breast Milk Relative to Dietary Contaminant Intake by the Mother

The units used for B_{bmk} are days/kg (mothers milk) (d/kg [mothers milk]).

No experimental results quantifying B_{bmk} are available in the current literature. An estimation method (Smith, 1987), is therefore considered:

Estimation Method

$$B_{\text{bmk}} = 2 \times 10^{-7} K_{\text{ow}}$$

$$K_{\text{ow}} = 28$$

$$B_{\text{bmk}} = 5.6 \times 10^{-6} \text{ days/kg (mothers' milk)}$$

The CV of the above method is approximately 10.

Based on the estimation equation and the estimation error reported above, we obtain the following value for the estimated biotransfer factor for human breast milk concentration relative to dietary contaminant intake by the mother for 1,2 DCA:

Arithmetic mean (coefficient of variation):
 $B_{\text{bmk}} = 5.6 \times 10^{-6}$ (10) days/kg (mothers' milk)

BCF: Bioconcentration Factors for Fish Relative to Water Concentration

The units used for BCF (fish/water) are $\frac{\text{mg/kg (fish)}}{\text{mg/liter (water)}}$, and BCF is therefore unitless.

Experimental Values:

2 reported at 16 °C as 2 BCF fish/water by Veith et al. (1980) using bluegill sunfish and a concentration of 96 µg/l

From the measured value* reported above, and the assumption that the variation in this measurement is approximately 1, we obtain the following statistics for BCF in fish relative to contaminant concentration in water for 1,2 DCA:

Arithmetic mean (coefficient of variation):
BCF (fish/water) = 2 (1)

*lipid-based BCFs are adjusted downward by a factor of 0.08 to account for fish lipid content

Estimation Method

For fish, the BCF is taken as the ratio of concentration of a xenobiotic substance in fish flesh (or lipids) to the contaminant's concentration in water (Mackay, 1982). The BCF for neutral organic compounds can be estimated from regression equations based on selected physicochemical properties, particularly a compound's K_{ow} or aqueous solubility. Mackay (1982) recommends:

$$\text{BCF} = 0.048 K_{ow}$$

$$K_{ow} = 28$$

$$\text{BCF (est)} = 1.3$$

$$\text{CV} = 0.6$$

The reported GSD is 1.8 which corresponds to an estimation error of 0.6.

 K_{p_w} : Human Skin Permeability Coefficient Relative to Contaminant Concentration in Water

The units used for K_{p_w} are centimeters/hour (cm/hr).

No reported measurements of K_{p_w} for 1,2 DCA are available in the current literature. An estimation method for this parameter is thus applied.

Estimation Method

Because dermal transfer is considered a nonsteady-state event, diffusion models require input parameters which are difficult to measure, such as the stratum corneum diffusion coefficient (D_{sc}) [Flynn and Amidon (1991)]. Estimation of aqueous biotransfer of 1,2 DCA is calculated with the following equation based on the estimation method of McKone and Howd (1992).

$$K_{p_w} = MW^{-0.6} 0.33 + \frac{0.0025}{2.4 \times 10^{-6} + 3 \times 10^{-5} K_{ow}^{0.8}}^{-1}$$

$$K_{ow} = 28$$

$$MW = 96.97 \text{ g/mol}$$

$$K_{p_w} = 0.010 \text{ cm/hr}$$

who report a coefficient of variation equal to 2.4

Based on the estimation equation and the estimation error reported above, we obtain the following value for the estimated human skin permeability coefficient relative to contaminant water concentration for 1,2 DCA:

Arithmetic mean (coefficient of variation):

$$K_{p_w} = 0.010 (2.4) \text{ cm/hr}$$

K_m : Partition Coefficient for Human Skin Relative to Contaminant Concentration in Water or Soil

The units used for K_m are $\frac{\text{mg/kg (skin)}}{\text{mg/liter (water)}}$ (ppm [skin]/ppm [water]).

No experimental values for K_m are currently available in the literature, therefore an estimation method is considered.

Estimation Method

Experimental values quantifying dermal transfer of 1,2 DCA in water, or for water in a soil matrix, may depend on pH, particle size and organic carbon content (Flynn and Amidon, 1991). An estimation method based on McKone and Howd (1992) is therefore used here.

$$K_m = 0.64 + (0.25 K_{ow}^{0.8})$$

$$K_{ow} = 28$$

$$K_m = 4.2 \text{ ppm (skin) / ppm (water)}$$

The reported geometric standard deviation of 1.3 in this estimation method corresponds to a CV of 0.27.

Based on the estimation equation and the estimation error reported above, we obtain the following value for the partition coefficient into human skin relative to 1,2 DCA water or soil concentration:

Arithmetic mean (coefficient of variation):
 $K_m = 4.2 (0.27) \text{ ppm (skin) / ppm (water)}$

T_{half_a}: Reaction Half-Life in Air

The units used for T_{half_a} are days.

Experimental Values

- 9.8 reported as a photo-oxidation half-life of 234 hours by Radding et al. (1977) [Callahan et al. (1979)]
- 36 reported at 23 °C as a photooxidation rate constant of $22 \times 10^{-14} \text{ cm}^3/\text{molecule-sec.}$ by Howard and Evenson (1976) using hydroxyl radicals and assuming a hydroxyl radical concentration of $1 \times 10^6 \text{ molecules/cm}^3$
- 12 to 122 reported as a photo-oxidation half-life of 292 to 2917 hours by Atkinson (1985) from the vapor phase reaction with hydroxyl radicals in air [also cited by Howard (1991) and MacKay et al. (1992)]

From the 4 experimental values reported above, we obtain the following statistics on the reaction half-life for 1,2 DCA in air:

Arithmetic mean (coefficient of variation):
 $T_{\text{half}_a} = 45 (1.2) \text{ days}$

Range: 9.8 to 122 days

T_{half_g}: Reaction Half-Life in Ground-Surface Soil

The units used for T_{half_g} are days.

Estimated Values

- 71 to 118 reported at 25 °C as an aerobic degradation rate in aquifer material of 15.12 and 23.85% by Watwood et al. (1991) using a Sandia and a Bosque unsaturated calcareous soils from New Mexico, 100 ppb, and ¹⁴C labelled 1,2 DCA for 4 weeks and assuming first-order degradation with respect to concentration
- 77 to 168 reported as a half-life of 11-24 weeks by Wilson et al. (1983) using soil from a shallow water-table aquifer at 20 °C for 16 weeks and assuming first-order degradation with respect to concentration [also cited in Howard (1991) and Mackay et al. (1992)]

From the 4 measured values above, we obtain the following statistics for the reaction half-life for 1,2 DCA in surface soil:

Arithmetic mean (coefficient of variation):

$$T_{\text{half}_g} = 110 (0.41) \text{ days}$$

Range: 71 to 168 days

T_{half_s}: Reaction Half-Life in Root-Zone Soil

The units used for T_{half_s} are days.

Estimated Values

- 71 to 118 reported at 25 °C as an aerobic degradation rate in aquifer material of 15.12 and 23.85% by Watwood et al. (1991) using a Sandia and a Bosque unsaturated calcareous soils from New Mexico, 100 ppb, and ¹⁴C labelled 1,2 DCA for 4 weeks and assuming first-order degradation with respect to concentration
- 77 to 168 reported as a half-life of 11-24 weeks by Wilson et al. (1983) using soil from a shallow water-table aquifer at 20 °C for 16 weeks and assuming first-order degradation with respect to concentration [also cited in Howard (1991) and Mackay et al. (1992)]

From the 4 measured values above, we obtain the following statistics for the reaction half-life for 1,2 DCA in root-zone soil:

Arithmetic mean (coefficient of variation):

$$T_{\text{half}_s} = 110 (0.41) \text{ days}$$

Range: 71 to 168 days

T_{half_v} : Reaction Half-Life in Vadose-Zone Soil

The units used for T_{half_v} are days.

Experimental Values

71 to 118 reported at 25 °C as an aerobic degradation rate in aquifer material of 15.12 and 23.85% by Watwood et al. (1991) using a Sandia and a Bosque unsaturated calcareous soils from New Mexico, 100 ppb, and ¹⁴C labelled 1,2 DCA for 4 weeks and assuming first-order degradation with respect to concentration

77 to 168 reported as a half-life of 11-24 weeks by Wilson et al. (1983) using soil from a shallow water-table aquifer at 20 °C for 16 weeks and assuming first-order degradation with respect to concentration [also cited in Howard (1991) and Mackay et al. (1992)]

568 to 573 reported at 25 °C as an anaerobic degradation rate in aquifer material of 3.33 and 3.36 % by Watwood et al. (1991) using a Sandia and a Bosque unsaturated calcareous soils from New Mexico, 100 ppb, and ¹⁴C labelled 1,2 DCA for 4 weeks and assuming first-order degradation with respect to concentration

From the 6 values above, we obtain the following statistics for the reaction half-life for 1,2 DCA in vadose-zone soil:

Arithmetic mean (coefficient of variation):

$$T_{\text{half}_v} = 260 (0.92) \text{ days}$$

Range: 71 to 573 days

T_{half_q}: Reaction Half-Life in Groundwater

The units used for T_{half_q} are days.

Experimental Values

- | | |
|---------------|--|
| 71 to
118 | reported at 25 °C as an aerobic degradation rate in aquifer material of 15.12 and 23.85% by Watwood et al. (1991) using a Sandia and a Bosque unsaturated calcareous soils from New Mexico, 100 ppb, and ¹⁴ C labelled 1,2 DCA for 4 weeks and assuming first-order degradation with respect to concentration |
| 77 to
168 | reported as a half-life of 11-24 weeks by Wilson et al. (1983) using soil from a shallow water-table aquifer at 20 °C for 16 weeks and assuming first-order degradation with respect to concentration [also cited in Howard (1991) and Mackay et al. (1992)] |
| 568 to
573 | reported at 25 °C as an anaerobic degradation rate in aquifer material of 3.33 and 3.36 % by Watwood et al. (1991) using a Sandia and a Bosque unsaturated calcareous soils from New Mexico, 100 ppb, and 14 C labelled 1,2 DCA for 4 weeks and assuming first-order degradation with respect to concentration |
| 289 | reported as a biotransformation rate constant of 1×10^{-4} hrs ⁻¹ by Silka (1988) using data from measurements at a sand and gravel outwash aquifer located in Tacoma, WA |

From the 7 values above, we obtain the following statistics for the reaction half-life for 1,2 DCA in groundwater:

Arithmetic mean (coefficient of variation):

$$T_{\text{half}_q} = 270 (0.83) \text{ days}$$

Range: 71 to 573 days

T_{half_w}: Reaction Half-Life in Surface Water

The units used for T_{half_w} are days.

Experimental Values

- | | |
|-----|--|
| 21 | reported at 25 °C as a half-life in water of 21 days by Tabak et al. (1981) using domestic wastewater conditions in a static-culture flask-screening procedure for 28 days in the dark |
| 540 | reported as a dark phase half-life in water of 18 months by Dilling (1975) assuming first order kinetics |

From the 2 values above, and our assumption that the range of actual values could be a factor of 5 higher or lower than this range, we obtain the following statistics for the half-life for 1,2 DCA in surface water:

Arithmetic mean (coefficient of variation):

$$T_{\text{half}_w} = 820 (1.6) \text{ days}$$

Range: 4 to 2700 days

T_{half_d}: Reaction Half-Life in Surface Water Sediment

The units used for T_{half_d} are days.

Experimental Values

- | | |
|---------------|--|
| 21 | reported at 25 °C as a half-life in water of 21 days by Tabak et al. (1981) using domestic wastewater conditions in a static-culture flask-screening procedure for 28 days in the dark |
| 540 | reported as a dark phase half-life in water of 18 months by Dilling (1975) assuming first order kinetics |
| 568 to
573 | reported at 25 °C as an anaerobic degradation rate in aquifer material of 3.33 and 3.36 % by Watwood et al. (1991) using a Sandia and a Bosque unsaturated calcareous soils from New Mexico, 100 ppb, and 14 C labelled 1,2 DCA for 4 weeks and assuming first-order degradation with respect to concentration |

From the 4 values above, we obtain the following statistics for the half-life of 1,2 DCA in sediment:

Arithmetic mean (coefficient of variation):

$$T_{\text{half}_d} = 430 (0.63) \text{ days}$$

Range: 21 to 573 days

References

- Ashworth, R.A., et al. 1986. Air-Water Partitioning Coefficients of Organics in Dilute Aqueous Solutions, paper presented at the *American Institute of Chemical Engineers 1986 National Meeting*, Boston, MA
- Atchison, J. and J.A.C. Brown, 1957. *The Lognormal Distribution*. Cambridge University, New York, pp. 125

- Atkinson, R., 1985. Kinetics and Mechanisms of the Gas Phase Reaction of Hydroxyl Radical with Organic Compounds Under Atmospheric Conditions, *Chem. Rev.* 85: 69–201
- Atkinson, R., 1989. Kinetics and Mechanisms of the Gas-Phase Reactions of the Hydroxyl Radical with Organic Compounds. *J. of Phy. and Chem. Ref. Data. Monograph No. 1*
- Banerjee, S., S.H. Yalkowsky, and S.C. Valvani 1980. Water Solubility and Octanol/Water Partition Coefficient of Organics. Limitations of Solubility-Partition Coefficient Correlation, *Environ. Sci. Technol.* 14(10):1227-1229
- Beratergremium Fur Umweltrelevante Altstoffe (BUA) Der Gesellschaft Deutscher Chemiker (Consulting Group Of The German Chemical Society For Environmentally Relevant Existing Chemicals), 1987. Dichlormethan BUA-Stoffbericht Nr. 6 (Report No. 6 on Dichloromethane). VCH New York, NY
- Boublik, T., V. Fried, and E. Hála, 1984. *The Vapour Pressures of Pure Substances: Selected Values of the Temperature Dependence of the Vapour Pressures of Some Pure Substances in the Normal and Low Pressure Region (Second Revised Edition)*, Physical Sciences Data; 17 Elsevier Science, Amsterdam, The Netherlands)
- Bruggeman, R., Trapp, S., Matthies, M., 1991. Behavior Assessment of a Volatile Chemical in the Rhine River. *Environ. Toxicol. & Chem.* 10:1097-1103
- Callahan, M.A., et al, 1979. Chapter 44. 1,2-Dichloroethane (Ethylene Dichloride). 2: Water-Related Environmental Fate of 129 Priority Pollutants. Vol. 2. EPA-440/4-79-029b. U.S. Environmental Protection Agency, Washington, DC 44-1 to 44-10
- Chiou, C.T., L.J. Peters, and A.H. Freed 1979. A Physical Concept of Soil-Water Equilibria for Nonionic Organic Compounds, *Science* 206:831–832
- Clayton, G. D. and Clayton, F. E. Ed., 1981. *Patty's Industrial Hygiene & Toxicology*. Third ed., John Wiley & Sons, New York. Vol. 2B:3491-3497
- Coca, J. and R. Diaz, 1980. *J. Chem. Eng. Data* 25(1):80-83
- Dean, J.D., Ed. 1985. *Lange's Handbook of Chemistry* McGraw-Hill, New York City, NY, 13th ed
- Dilling, W. L. 1977. *Environ. Sci. Technol.* 11(4):405-409
- Dilling, W.L., N.B. Tefertiller and G.J. Kallos, 1975. Evaporation Rates and Reactivities of Methylene Chloride, Chloroform, 1,1,1-Trichloroethane, Trichloroethylene, Tetrachloroethylene, and Other Chlorinated Compounds in Dilute Aqueous Solutions. *Environ. Sci. Technol.* 9(9):833-838
- EPA/OTS, 1987. *Analysis and Interpretation of Batch Equilibrium and Column Studies of the Partitioning of Chlorinated Hydrocarbons to Soil Materials from the PPG Lake Charles Site*, Doc #86-870002008,
- Flynn and Amidon, 1991. *Dermal Exposure Assessment: Principles and Applications*, Interim Report. USEPA Jan, 1992. Office of Research and Development. Washington, DC. EPA/600/8-91/011B
- Fuller, E.N., P.D. Schettler, and J.C. Giddings, 1966. A New Method for Prediction of Binary Gas-Phase Diffusion Coefficients, *Ind. Eng. Chem.* 58:19-27

Gross, P. M., 1929. The Determination of the Solubility of Slightly Soluble Liquids in Water and the Solubilities of Dichloroethanes and Dichloropropanes, *J. Am. Chem. Soc.* 51:2362-2366

Gross, P.M., and J.H. Saylor, 1931. The Solubilities of Certain Slightly Soluble Organic Compounds in Water, *J. Am. Chem. Soc.* 53:1744-1751

Gutsche, B., and H., Knapp, 1982. *Fluid Phase Equilibria* 8:285

Hamburg, M. 1970. *Statistical Analysis for Decision Making*. Harcourt Brace Jovanovich; San Diego, California

Hansch, C., and A. Leo 1987. The Log P and Related Parameters Database, created and updated by the Medicinal Chemistry Project at Pomona College, Claremont, CA, under the direction of Corwin Hansch and Albert Leo — 1987 Printout

Hansch, C., and A. Leo, 1985. The Log P and Related Parameters Database, created and updated by the Medicinal Chemistry Project at Pomona College under the direction of Corwin Hansch and Albert Leo — 1985 Printout, *Medchem Project Issue N^o. 26*, Pomona College, Claremont, CA

Hendry and Kenley, 1976. *Atmospheric Reaction Products of Organic Compounds*, EPA Report. EPA-560/12-79-001, 81 pp

Horvath, A. L., 1982. *Halogenated Hydrocarbons, Solubility-Miscibility with Water* Marcel Dekker, Inc., New York, NY

Howard, C. J. and Evenson, K. M., 1976. Rate Constants for the Reactions of OH with Ethane and Some Halogen Substituted Ethanes at 296 K, *J Chem. Phys.* 64(11):4303-4306

Howard, P. H., R. S. Boethling, W. F. Jarvis, W. M. Meylan, and E. M. Michalenko, Eds. 1991. *Handbook of Environmental Degradation Rates* Lewis; Chelsea, MI

Howard, P.H., G.W. Sage, W.F. Jarvis, and D.A. Gray, Eds. 1990. *Handbook of Environmental Fate and Exposure Data for Organic Chemicals. Volume II. Solvents* Lewis; Chelsea, MI

Jow, P. and Hansch, 1985. Pomona College, Unpublished Results

Karickhoff, S. W., 1981. Semi-Empirical Estimation of Sorption of Hydrophobic Pollutants of Natural Water Sediments and Soils, *Chemosphere* 10(8):833-846

Kenaga, E. E., 1980. Correlation of Bioconcentration Factors of Chemicals in Aquatic and Terrestrial Organisms with Their Physical and Chemical Properties. *Environ. Sci. Technol.*, 14(5):553-556

Kirk-Othmer Encyclopedia of Chemical Technology, 1964. *Chlorocarbons and Chlorohydrocarbons*, Vol. 5, 2nd ed., Wiley Interscience; New York, NY. pp 100-363

Liljestrand, H. M. and R. J. Charbeneau, 1987. Analysis and Interpretation of Batch Equilibrium and Column Studies of the Partitioning of Chlorinated Hydrocarbons to Soil Materials from the PPG Lake Charles Site. Environmental and Water Resources Engineering, Department of Civil Engineering, The University of Texas at Austin. Austin, TX 78712

Lyman, W. J., W. F. Reehl, and D. H. Rosenblatt, 1990. *Handbook of Chemical Property Estimation Methods: Environmental Behavior of Organic Compounds*, American Chemical Society, Washington, DC

Lyman, W. J., W. F. Reehl, and D. H. Rosenblatt, 1982. *Handbook of Chemical Property Estimation Methods: Environmental Behavior of Organic Compounds*. American Chemical Society, Washington, D. C.

Mackay, D. 1982. Correlation of Bioconcentration Factors, *Environ. Sci. Technol.* 16:274-278

Mackay, D., W.-Y. Shiu, and K.-C. Ma, 1993. *Illustrated Handbook of Physical-Chemical Properties and Environmental Fate for Organic Chemicals. Volume III. Volatile Organic Chemicals*, Lewis; Boca Raton, FL

McGovern, E. W., 1943. Chlorohydrocarbon Solvents. *Ind. Eng. Chem.* 35(12):1230-1239

McKone, T. E., and Howd, R. A., 1992. Estimation Dermal Uptake of Nonionic Organic Chemicals from Water and Soil: I. Unified Fugacity-Based Models for Risk Assessments. *Risk Analysis* 12(4):543-557

McKone, T. E., and Ryan, P. B., 1989. Human Exposures to Chemicals Through Food Chains: An Uncertainty Analysis. *Environ. Sci. Technol.* 23:1154-1163

McKone, T.E., 1993. The Precision of QSAR Methods for Estimating Intermedia Transfer Factors in Exposure Assessments, *SAR and QSAR in Environmental Research*, 1:41-51

McNally, M.E., and R.L. Grob, 1984. Headspace Determination of Solubility Limits of the Base Neutral and Volatile Components from Environmental Protection Agency's List of Priority Pollutants. *J. Chromatog.* 284:105-116

Morgan, D. L., Cooper, S W., Carlock, D L., Sykora, J J., Sutton, B., Mattie, DR., McDougal, JN. Dermal Absorption of Neat and Aqueous Volatile Organic Chemicals in the Fischer 344 Rat. *Environ-Res*, Vol 55(1):51-63

Mudder, T. 1981. Development of Empirical Structure-Biodegradability Relationships and Testing Protocol for Slightly Soluble and Volatile Priority Pollutants, *Diss. Abstr. Int. B.* 42:1804

Pearce, J. N., and P. E. Peters, 1929. *J. Phys. Chem.* 33:873

Radding, S. B., Lui, D. H., Johnson, H. L., & Mill, T. 1977. Review of the Environmental Fate of Selected Chemicals, Washington, DC, US EPA, office of Toxic Substances. EPA 560/5-77-003, PB 267121

Reid, R. C., J. M. Prausnitz, and B. E. Poling, 1987. *The Properties of Gases and Liquids* McGraw Hill, New York, NY 4th ed

Reiderer, M. 1990. "Estimating Partitioning and Transport of Organic Chemicals in the Foliage/Atmosphere System: Discussion of a Fugacity-Based Model," *Environ. Scil. Technol.* 24:829-837

Riddick, J. A. , Bunger, W. B., Sakano, T. K., 1986. *Organic Solvents: Physical Properties and Methods of Purification*. 4th ed., John Wiley & Sons. New York, NY Vol. II: No. 319

- Scherb, K., 1978. [Investigations on the Evaporation of Low Molecular Weight Chlorinated Hydrocarbons from Flow-Gutter] *Munch. Beitr. Abwasser-Fisch. Flussbiol.*, 30:235-248 (in German)
- Schwarzenbach, R. P., Gschwend, P. M., and D. M. Imboden, 1993. *Environmental Organic Chemistry*. Wiley-Interscience, John Wiley and Sons, New York, NY
- Silka, L.R., and D.A. Wallen, 1988. *Observed Rates of Biotransformation of Chlorinated Aliphatics in Groundwater*. Superfund '88, Proceedings of the 9th National Conference, Nov. 28-30, 1988, Washington, DC Hazardous Materials Control Research Institute, Washington, DC pp. 138-141
- Smith, A. H. 1987. Infant Exposure Assessment for Breast Milk Dioxins and Furans Derived from Waste Incineration Emissions. *Risk Analysis* 7:347-353
- Stephens, R. D., Harnly, M., Haywood, G. G., Chang, R. R., Flattery, J., Petreas, M. X., Goldman, L., 1990. Bioaccumulation of Dioxins in Food Animals II: Controlled Exposure Studies, *Chemosphere* 20:1091-1096
- Stull, D. R. 1947. Vapor Pressure of Pure Substances: Organic Compounds, *Ind. Eng. Chem.* 39:517-540
- Tabak, H. H., Quave, S. A., Mashni, C. I., and Barth, E. F., 1981. Biodegradability Studies with Organic Priority Pollutant Compounds. *J. Water Pollut. Control Fed.*, 53:153-1518
- Thermodynamics Research Center Data Project, Thermodynamics Research Center, College Station, Texas A&M University. Table 23-10-2-(2.013)-a, Dec. 31, 1964
- Travis, C. C., and A. D. Arms, 1988. Bioconcentration of Organics in Beef, Milk, and Vegetation, *Environ. Sci. Technol.* 22:271-274
- Tse, G., Orbey, H., and Sandler, S. I. Infinite Dilution Activity Coefficients and Henry's Law Coefficients of Some Priority Water Pollutants Determined By a Relative Gas Chromatographic Method. *Environmental Science & Technology* 26(10):2017-2022
- US Department of Health and Human Services, Toxicological Profile for 1,2 Dichloroethane. Draft, 1993
- Veith, G.D., K.J. Macek, S.R. Petrocelli, and J. Carroll 1980. An Evaluation of Using Partition Coefficients and Water Solubility to Estimate Bioconcentration Factors for Organic Chemicals in Fish, in *Aquatic Toxicology*, ASTM STP 707, J. G. Eaton, P. R. Parrish, and A. C. Hendricks, Eds. American Society for Testing and Materials, Philadelphia, PA pp 117-119
- Verschuere, K., 1983. *Handbook of Environmental Data on Organic Chemicals*, Second Edition, Van Nostrand Reinhold, New York, NY
- Warner, H.P., J.M. Cohen and J.C. Ireland, 1987. *Determination of Henry's Law Constants of Selected Priority Pollutants.*, EPA/600/D-87/229 U.S. EPA, Cincinnati, OH. NTIS: PB87-212684 14
- Weast, R. C., Lide, D. R., Astle, M.J. and Beyer, W.H. Eds. 1989. *CRC Handbook of Chemistry and Physics: A Ready Reference Book of Chemical and Physical Data*, 67th ed. CRC Press, Boca Raton, FL
- Wilke, C.R., and P. Chang, 1955. Correlation of Diffusion Coefficients in Dilute Solutions, *AIChE J.* 1:264-270

Wilson, J.T., McNabb, J. F., Blackwill, D.L., Ghiorse, W. C., 1983. Enumeration and Characterization of Bacteria Indigenous to a Shallow Water-table Aquifer. *Ground Water* 21:134-142.

World Health Organization International Agency for Research on Cancer, 1985. Monographs on the Evaluation of the Carcinogenic Risk of Chemicals to Humans: Some Chemicals Used in Plastics and Elastomers, 39:195-215

Wright, W.H., and J.M. Schaffer, 1932. Critical Anthelmintic Tests of Chlorinated Alkyl Hydrocarbons and a Correlation Between the Anthelmintic Efficacy, Chemical Structure and Physical Properties, *Am. J. Hyg.* 16(2):325-428